

## Influence of Light Intensity on the Photosynthesis and Phenolic Contents of *Mangifera Indica*

Alona C. Linatoc<sup>1</sup> and Aisha Idris<sup>1,2\*</sup> Mohd Fadzelly Abu Bakar<sup>1</sup>

<sup>1</sup>Faculty of Applied Sciences and Technology, Universiti Tun Hussein Onn Malaysia (UTHM), Hub Pendidikan Tinggi Pagoh, KM1, Jalan Panchor, 84600, Muar, Johor, Malaysia

<sup>2</sup>Faculty of Sciences, Federal University Dutse, PMB 7156, Jigawa State, Nigeria

Received 15 September 2018; accepted 1 December 2018; available online 30 December 2018

DOI: <https://10.30880/jst.2018.10.04.009>

**Abstract:** Light is an important environmental factor that have an influence on a plants photosynthesis and production of secondary metabolites like phenolic compounds and flavonoid. *Mangifera indica* from the family *Anacardiaceae* is known to have bioactivity due to its phenolic and flavonoid contents. The objective of the study is to determine the influence of light on the photosynthesis and phenolic contents of *M. indica*. Photosynthesis of the plant was measured using a portable photosynthesis system referred to as LICOR- 6400. Photosynthetic pigments as well as phenolic and flavonoid contents were quantified using a UV-VIS spectrophotometer. The outcome derived from the study shows that sun exposed leaves of the studied plant were having the maximum photosynthesis, saturation and compensation points ( $P < 0.05$ ). Moreover, sun exposed leaves were having higher carotenoid, phenolic and flavonoid contents but lower chlorophyll contents. This leads to a conclusion that sun leaves of *M. indica* contribute the highest photosynthesis and phenolic contents to the plant.

**Keyword:** Flavonoid, light intensity; *Mangifera indica*; phenolic; photosynthetic pigment.

### 1.0 Introduction

*Mangifera Indica* (Mango) is a fruit plant belonging to the *Anacardiaceae* family. It is widely distributed in the world, even though it is native to Indian subcontinent. Mango is well known to contain some active ingredient that contribute to its traditional medicine application. Shah et al. [1] reported the use of mango tree and its part as food and as drug. Different part of the tree can be extracted and use as antibiotic. Other uses of the tree include the utilization of the wood to make lumber which can be used to make ukuleles and furniture [2]. Studies on mango are mainly concerned about their nutritional [3], antioxidant, antiviral, antidiabetic, anthelmintic, ant parasitic, antidiarrheal, and anti-inflammatory [4][5][3][1][6][7][5][8][9]. Moreover, *M. indica* can be used as a probiotic ingredient [3].

Environmental factors like light play an important role in the accumulation of phenolics in plants. This is due to the fact that phenolics are produced in response to adverse conditions, like insect attack, UV radiation, as well as drought and parasite attack. Moreover, biomass accumulated by a particular plant is also

affected by the level of light intensity received by the plant [10]. The photosynthesis of a plant depends on the light received by the plants [11], due to this the pigment content of a particular plant changes depending on the level of light available to the plant. For example, Chl a, Chl b and Carotenoid contents of *Lactuca sativa* was highest when the plant was grown under blue LED light while Chl a/b ratio was highest under red LED light [12]. Because light intensity affects a plants photosynthesis, the net assimilation rate ( $A_{net}$ ) of *L. sativa* was highest under red light compared to blue LED light [12].

Phenolic contents of *Lactuca sativa* was highest when the plant was grown under white LED light [12]. Another study by Ballester et al. [13] revealed that blue LED light do not influence the phenolic contents of fruits. Total phenolic content of *Brassica oleracea* was highest under red LED [14]. In addition to this, *Zea mays* sprouts that germinate under light have more phenolic content than those that germinate under dark conditions. This is also true for the flavonoid content of the plant [15].

The objective of this research is to determine the influence of varying light

\*Corresponding author: [hw160077@siswa.uthm.edu.my](mailto:hw160077@siswa.uthm.edu.my)

2018 UTHM Publisher. All right reserved.

e-ISSN: 2600-7924/[penerbit.uthm.edu.my/ojs/index.php/jst](http://penerbit.uthm.edu.my/ojs/index.php/jst)

intensity on the photosynthesis and phenolic contents of *Mangifera indica*.

## 2. Materials and method

### 2.1 Determination of the effect of light of the plant photosynthesis

Sun and shade leaves of *M. indica* were used for this study. Prior to the study, a branch of sun and shade leaves were cut out of the plant, inserted into a flask of water, recut under water, and then incubated at ambient conditions for 24 hours. After the incubation, the portable photosynthesis system was assembled and set to the desired parameters, where flow rate was  $500 \mu\text{mol m}^{-2} \text{s}^{-1}$ ,  $\text{CO}_2$  and relative humidity were ambient while photosynthetic active radiation was variable from  $2000 \mu\text{mol m}^{-2} \text{s}^{-1}$  to  $0 \mu\text{mol m}^{-2} \text{s}^{-1}$ . Readings were recorded and then used to construct a photosynthetic light response curve. Moreover, parameters like  $A_{\text{net}}$ , maximum photosynthesis ( $A_{\text{max}}$ ), apparent quantum yield ( $A_{\text{qy}}$ ), light saturation point (LSP) and light compensation points (LCP) were deducted after fitting the curve according to Marshall & Biscoe [16].

### 2.2 Determination of the photosynthetic pigment contents

Photosynthetic pigments were determined spectrophotometrically were leaf sample were suspended in ethanol and quantified according to Lichtenthaler & Buschmann [17] after recording the absorbance of Chl a at 664.2nm, Chl b at 648.6 nm and carotenoid at 470nm respectively.

### 2.3 Plant material

Sun exposed and shaded mango tree leaves were sampled and harvested. 100g was measured and dried in an oven at  $45^\circ\text{C}$  for 42 hours. Sample was extracted based on Chai et al. [18] by grinding the dried leaves into powder. Furthermore, the dried powder was mixed with 2000 ml of ethanol and then incubated for 2 hours. The plant mixture was then filtered with a Whatman filter paper prior to centrifuging at 12000 rpm, at  $4^\circ\text{C}$  for 10 minutes. The supernatant was stored at low temperature and later used for determining total flavonoid and phenolic contents of the plants.

### 2.4 Determination of the total flavonoid contents

Total flavonoid content of *M. indica* was quantified using UV-vis spectrophotometer. The procedure was based on the aluminium chloride calorimetry method [6]. Plant extract was prepared and reaction mixture was developed by mixing plant extract with  $\text{AlCl}_3$  and potassium acetate. Absorbance was recorded at 420 nm. Calibration curve was generated using Quercetin. Furthermore, results were expressed as mg/g quercetin equivalents (QE).

### 2.5 Determination of the total phenolic contents

Total phenolic content present in *M. indica* were quantified using UV-vis spectrophotometer [19]. The procedure was based on the folin-chiocalteau reagent. Quantification was done using UV-visible spectrophotometer. The leaf extract was first extracted and then the reaction mixture was developed using the reagent and  $\text{NaCO}_3$ . Absorbance was recorded at 760 nm, Gallic acid calibration curve generated, and results were expressed as mg/g gallic acid equivalents (GAE).

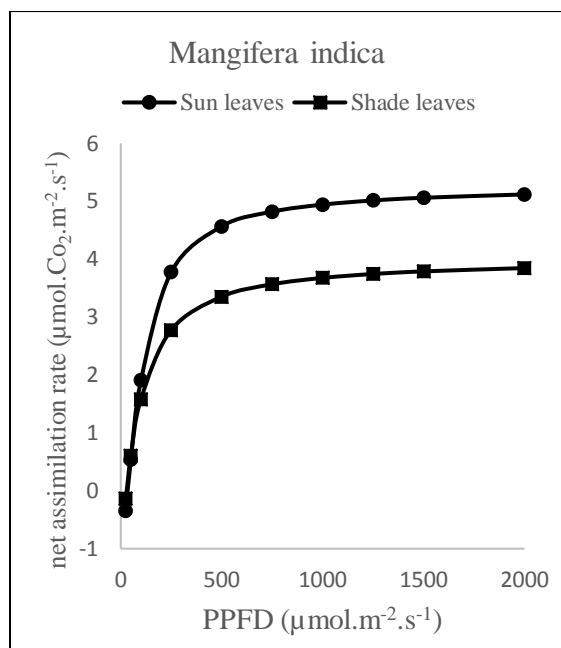
### 2.6 Statistical analysis

Data were compiled and recorded in triplicates. Results were reported as means  $\pm$  standard deviation of the means. Additionally, differences in mean of sun and shade leaves were compared using T-test. Statistical analysis was done using the SPSS statistical software (IBM corp).

## 3. Results and Discussion

### 3.2 Influence of light on the plant photosynthesis

Varying light intensity influences the assimilation rate of *M. indica*. Sun exposed leaves contribute to the highest photosynthesis of the plant. The maximum photosynthesis, LCP as well as the LSP were higher in sun leaves compared to shade leaves. In addition, the photosynthetic light response curve was higher in sun leaves compared to shade leaves (Fig. 1).



**Fig. 1** Photosynthetic light response curve of *M. indica*.

The maximum photosynthesis recorded in sun leaves is due to higher number of chloroplast usually present in sun leaves [20]. Even though the quantum yield of sun and shade leaves were not significantly different ( $P > 0.05$ ), the photosynthetic capacity of sun leaves is higher than that of shaded leaves. This is possible because the capacity is determined by the LSP and LCP of the leaves (Table 1).

**Table 1** Photosynthetic light response parameters of *M. indica*.

	$A_{max}$	LCP	LSP	$A_{qy}$
S	$6.62 \pm 0.12^a$	$34.30 \pm 2.1^a$	$976.73 \pm 4.213^a$	$0.042 \pm 0.009^a$
SH	$5.15 \pm 0.09^b$	$19.12 \pm 1.1^b$	$537.16 \pm 3.162^b$	$0.045 \pm 0.011^a$

Where  $A_{max}$  is maximum photosynthesis in  $\mu\text{mol.CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ , LCP is light compensation point in  $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ , LSP is light saturation point in  $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ ,  $A_{qy}$  is apparent quantum yield in  $\text{mol CO}_2 \text{ mol}^{-1} \text{ photons}$ , S is sun exposed leaves, and SH is shaded leaves. Different small letters indicate significant differences between sun and shade leaves ( $P < 0.05$ ).

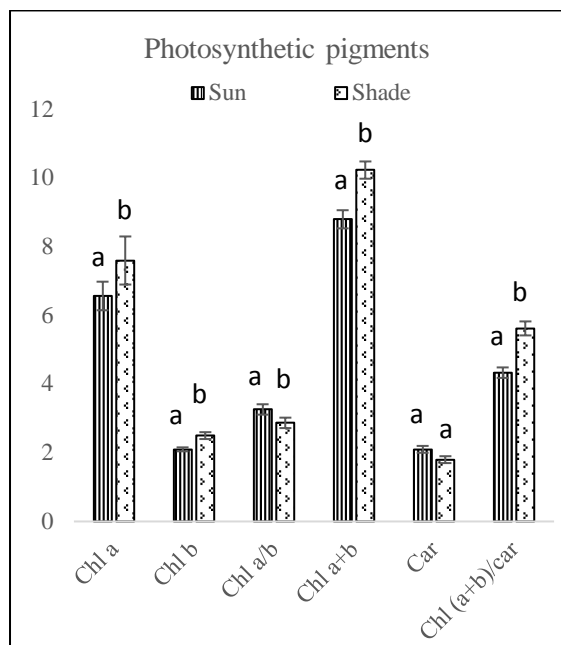
Leaves with lower LCP can use lower light intensity more than leaves with higher compensation point. The light intensity needed by a leaf to reach its maximum photosynthesis is referred to as the LSP. Above the LSP of a leaf, light will no longer be the limiting factor for photosynthesis, but rather, other factors like carboxylation rate will be the determining factors. Below the LSP, light will be the limiting factor for photosynthesis [20]. In a study, a plastic roof was used to provide shading to mango leading to a reduction in the photosynthetic photon flux density (PPFD) by about  $700 \mu\text{mol m}^{-2} \text{ s}^{-1}$ . This leads to an increase in the  $A_{net}$  of the plant [21]. The maximum photosynthesis of *M. indica* obtained in this study is  $6.62 \pm 0.12$  for sun leaves. In another study, the photosynthesis of *M. indica* ranges between 4 to  $11 \mu\text{mol m}^{-2} \text{ s}^{-1}$  [22]. Light was reported to affect the photosynthesis of *Arabidopsis thaliana* [23], while far red light positively influence the photosynthesis *Lactuca sativa* [24].

In *Camptotheca acuminata* [25], red light can increase the plants biomass, leaf area, specific leaf area, Chl a and b, as well as carotenoid contents. Besides,  $A_{net}$ , stomatal conductance, and transpiration rate also increases under red light treatment. Moreover, chloroplast width, granular number and thickness, as well as number of grana lamellae were higher under red light compared to white, blue and yellow LED light [25]. Furthermore, the photosynthetic light response curve of grape vines grown under different LED light was highest under blue light [26].

### 3.2 Influence of light on the plant photosynthesis pigments

The photosynthetic pigments of sun and shade leaves of *M. indica* is represented in Fig. 2. From the results obtained, it is obvious that shade leaves were having high number of Chl a and Chl b compared to sun leaves. Shade leaves are thinner than sun leaves, giving them an opportunity to maximize capturing light photons. Shade leaves were greener than sun leaves because they have higher ratio of total chlorophyll to carotenoid content. Moreover, because shade leaves have higher Chl b content, their chlorophyll a to b ratio is lower than that of sun leaves. Nevertheless, sun leaves had higher Chl a to b ratio because they adapt to light more than shade leaves. In addition, sun

exposed leaves had higher carotenoid content compared to shaded leaves due to the photo protective role of carotenoid. Carotenoid content of mango differs during the fruit development stage [27].



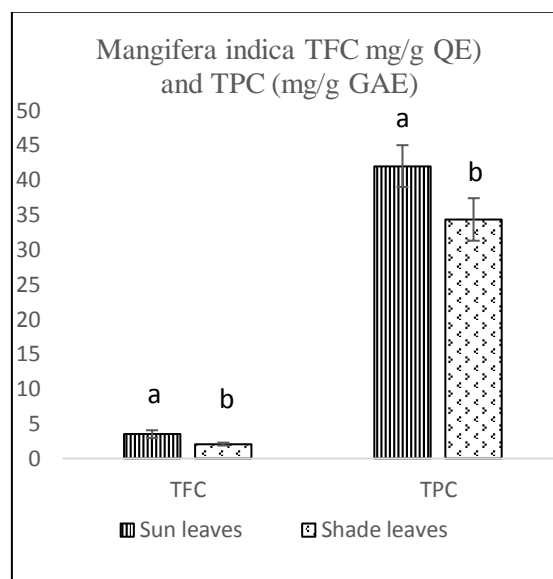
**Fig. 2** Photosynthetic pigments of *M. indica*

Subbaiah et al. [28] reported that the Chl a content of mango was about 2.13mg/g, while the Chl b content was about 0.64 mg/g. The carotenoid and chlorophyll content of mango were highest under high light intensity and lowest when the light intensity was reduced with yellow and black fruit bag [29]. Even the  $\beta$  carotene present in mango differs according to the light available. Ripped mango fruit contain the highest  $\beta$  carotene [30]. In *Brassica Campestris*, shade treatment can increase the Chl b content [31]. Moreover, the Chl content of *Nannochloropsis* species was highest under red light and lowest under white light [32].

### 3.2 Influence of light on the plant total phenolic and flavonoid contents

Phenolic and flavonoids contents present in *M. indica* were higher in sun exposed leaves compared to shaded leaves. In Fig. 3, it is seen that the differences between the phenolic and flavonoid contents of sun exposed and shaded leaves is statistically significant ( $P < 0.05$ ). Due to the fact that phenolics can accumulate in leaf epidermis as a result of adverse environmental conditions [33], *M. indica* sun exposed leaves accumulate higher amount of phenolics. Light

can affect the flavonoid concentration of plants, for example [34] reviewed the effect of light on the accumulation of flavonoids in *Ginkgo biloba*.



**Fig. 3** Total phenolic and flavonoid contents of *M. indica*

Different small letters indicate significant differences among sun and shade leaves ( $p < 0.05$ ).

In a recent study by De-Ancos et al. [35], the authors reported that the total phenolic content of mango was about 43 g/kg. Agatonovic-Kustrin et al. [36] on the other hand extracted about 26.92 mg/g GAE of phenolic contents in mango peel. In another study, the phenolic contents extracted from mango was about 57.86  $\mu\text{g}/\text{mg}$  [37]. Moreover, total phenolic content of mango extracted by Dorta et al. [38] ranges between 47 to 79 g/100g DW while the total flavonoid contents ranges from 3 to 33 g/100g. In addition, total phenolic contents extracted from mango was about 960.4 mg/kg GAE [39].

Blue light can influence the accumulation of phenolic content of *Lachenalia* species [19], while red light can influence the phenolic content of *Brassica oleracea* [14]. UV light affect the phenolics and pigment content of broccoli sprouts [40], carrots [41], *Vigna radiate* sprouts [42], *Gracilaria chilensis* [43], *Capsicum chinense* [44], and *Solanum lycopersicum* [45]. High light intensity affects the phenolic and flavonoid content of *Salvia plebeia* [46], *Brassica* species [47] and *L. sativa* [48]. Even mulberry treated with pulsed light

ultrasound have higher phenolic content than mulberry treated with other ultrasound treatments [49].

#### 4.0 Conclusion

To this point, it is obvious that the photosynthesis, pigment and phenolic content of *M. indica* depends on the available light. Furthermore, sun leaves contribute to the highest photosynthesis of the studied plant. Besides, the photosynthetic pigments of the plant varies, with higher concentration of Chl in shaded leaves and higher concentration of carotenoid in sun exposed leaves. In addition, the phenolic content of the studied plant are higher in sun exposed leaves compared to shaded leaves.

#### Acknowledgements

The authors gratefully acknowledge the Universiti Tun Hussein Onn Malaysia (UTHM) for the facilities provided.

#### REFERENCES

- [1] Shah, K. A., Patel, M. B., Patel, R. J., & Parmar, P. K. (2010). *Mangifera indica* (mango)., *Pharmacogn. Rev.*, vol. 4, no. 7, pp. 42–8.
- [2] Hardegee, M. C., & Tu, A. T. (1988). *Handbook of Natural Toxins: Bacterial Toxins*. CRC Press.
- [3] Hu, K., Dars, A. G., Liu, Q., Xie, B., & Sun, Z. (2018). Phytochemical profiling of the ripening of Chinese mango (*Mangifera indica* L.) cultivars by real-time monitoring using UPLC-ESI-QTOF-MS and its potential benefits as prebiotic ingredients, *Food Chem.*, vol. 256, no. 2017, pp. 171–180.
- [4] Mutua, J. K., Imathiu, S., & Owino, W. (2017). Evaluation of the proximate composition, antioxidant potential, and antimicrobial activity of mango seed kernel extracts, *Food Sci. Nutr.*, vol. 5, no. 2, pp. 349–357.
- [5] Irondi, E. A., Awoyale, W., Oboh, G., & Boligon, A. A. (2017). Effect of mango kernel flour addition on the phenolics profile, antioxidant activity and pasting properties of wheat flour, *J. Food Meas. Charact.*, vol. 11, no. 4, pp. 2202–2210.
- [6] Taj, I. K., Muhammad, N., Muhammad, I., Muhammad, A., & Sadaqat, A. (2018). Antioxidant activity, fatty acids characterization and oxidative stability of Gouda cheese fortified with mango (*Mangifera indica* L.) kernel fat, *J. Food Sci. Technol.*, vol. 55, no. 3, pp. 992–1002.
- [7] Anila, L., & Vijayalakshmi, N. R. (2003). Antioxidant action of flavonoids from *Mangifera indica* and *Emblica officinalis* in hypercholesterolemic rats, *Food Chem.*, vol. 83, no. 4, pp. 569–574.
- [8] Fessard, A., Kapoor, A., Patche, J., Assemat, S., Hoarau, M., Bourdon, E., & Remize, F. (2017). Lactic Fermentation as an Efficient Tool to Enhance the Antioxidant Activity of Tropical Fruit Juices and Teas, *Microorganisms*, vol. 5, no. 2, p. 23.
- [9] Jincy, M., Djanaguiraman, M., Jeyakumar, P., Subramanian, K. S., Jayasankar, S., & Paliyath, G. (2017). Inhibition of phospholipase D enzyme activity through hexanal leads to delayed mango (*Mangifera indica* L.) fruit ripening through changes in oxidants and antioxidant enzymes activity, *Sci. Hort. (Amsterdam)*, vol. 218, pp. 316–325.
- [10] Kurniawati, F. N., Mahajoeno, E., Sunarto, & Sari, S. L. A. (2017). Effect of Light Intensity for Optimum Biomass and Lipid Production from *Scenedesmus dimorphus* (Turpin) Kützing, *Int. Conf. Green Renew. Energy Resour.*, vol. 75, pp. 1–6.
- [11] Kim, J. H., Lee, J. W., Ahn, T. I., Shin, J. H., Park, K. S., & Son, J. E. (2016). Sweet Pepper (*Capsicum annuum* L.) Canopy Photosynthesis Modeling Using 3D Plant Architecture and Light Ray-Tracing, *Front. Plant Sci.*, vol. 7, no. September, pp. 1–10.
- [12] Amoozgar, A., Mohammadi, A., & Sabzalian, M. R. (2017). Impact of light-emitting diode irradiation on photosynthesis, phytochemical composition and mineral element content of lettuce cv. Grizzly, *Photosynthetica*, vol. 55, no. 1, pp. 85–95.
- [13] Ballester, A. R., & Lafuente, M. T. (2017). LED Blue Light-induced changes in phenolics and ethylene in citrus fruit: Implication in elicited

- resistance against *Penicillium digitatum* infection, *Food Chem.*, vol. 218, pp. 575–583.
- [14] Deng, M., Qian, H., Chen, L., Sun, B., Chang, J., Miao, H., & Wang, Q. (2017). Influence of pre-harvest red light irradiation on main phytochemicals and antioxidant activity of Chinese kale sprouts, *Food Chem.*, vol. 222, pp. 1–5.
- [15] Xiang, N., Guo, X., Liu, F., Li, Q., Hu, J., & Brennan, C. S. (2017). Effect of light- and dark-germination on the phenolic biosynthesis, phytochemical profiles, and antioxidant activities in sweet corn (*Zea mays* L.) sprouts, *Int. J. Mol. Sci.*, vol. 18, no. 6.
- [16] Marshall, B., & Biscoe, P. V. (1980). A Model for C 3 Leaves Describing the Dependence of Net Photosynthesis on Irradiance, *J. Exp. Bot.*, vol. 31, no. 1, pp. 29–39.
- [17] Lichtenthaler, H. K., & Buschmann, C. (2001). Chlorophylls and Carotenoids : Measurement and Characterization by UV-VIS, in *Current Protocols in Food Analytical Chemistry*, pp. 1–8.
- [18] Chai, T., Elamparuthi, S., Yong, A., Quah, Y., & Ong, H. (2013). Activities of selected highland ferns of Malaysia, *Botanical Studies*, vol. 54, no. 55, pp. 1–7.
- [19] Bach, A., Kapczyńska, A., Dziurka, K., & Dziurka, M. (2018). The importance of applied light quality on the process of shoot organogenesis and production of phenolics and carbohydrates in *Lachenalia* sp. cultures in vitro, *South African J. Bot.*, vol. 114, pp. 14–19.
- [20] Lambers, H., Chapin, F. S., & Pons, T. L. (2008). *Plant Physiological Ecology*, vol. 53, no. 9.
- [21] Juntamanee, K., Onnom, S., Yingjajaval, S., & Sangchote, S. (2013). Leaf photosynthesis and fruit quality of mango growing under field or plastic roof condition, *Acta Hort.*, vol. 975, no. 2, pp. 415–420.
- [22] Santos, M. R. dos, Martinez, M. A., Donato, S. L. R., & Coelho, E. F. (2014). Tommy Atkins' mango yield and photosynthesis under water deficit in semiarid region of Bahia, *Rev. Bras. Eng. Agrícola e Ambient.*, vol. 18, no. 9, pp. 899–907.
- [23] Köhl, K., Tohge, T., & Schöttler, M. A. (2017). Performance of *Arabidopsis thaliana* under different light qualities: Comparison of light-emitting diodes to fluorescent lamp, *Funct. Plant Biol.*, vol. 44, no. 7, pp. 727–738.
- [24] Zhen, S., & van Iersel, M. W. (2017). Far-red light is needed for efficient photochemistry and photosynthesis, *J. Plant Physiol.*, vol. 209, pp. 115–122.
- [25] Yu, W., Liu, Y., Song, L., Jacobs, D. F., Du, X., Ying, Y., & Wu, J. (2017). Effect of Differential Light Quality on Morphology, Photosynthesis, and Antioxidant Enzyme Activity in *Camptotheca acuminata* Seedlings, *J. Plant Growth Regul.*, vol. 36, no. 1, pp. 148–160.
- [26] Li, C. X., Chang, S. X., Khalil-Ur-Rehman, M., Xu, Z. G., & Tao, J. M. (2017). Effect of irradiating the leaf abaxial surface with supplemental light-emitting diode lights on grape photosynthesis, *Aust. J. Grape Wine Res.*, vol. 23, no. 1, pp. 58–65.
- [27] Ma, X., Zheng, B., Ma, Y., Xu, W., Wu, H., & Wang, S. (2018). Carotenoid accumulation and expression of carotenoid biosynthesis genes in mango flesh during fruit development and ripening, *Sci. Hortic. (Amsterdam)*, vol. 237, no. April, pp. 201–206.
- [28] Subbaiah, K. V., Reddy, N. N., Reddy, M. L. N., & Dorajeero, A. V. D. (2017). Effect of Different Irrigation Levels on Yield and Physiological-Biochemical Characteristics of Mango cv . Banganpalli, vol. 5, no. 6, pp. 177–182.
- [29] Ling, M. A. Z., & Bin, W. E. I. C. (2018). The Effect of Different Bags on Chlorophyll and Carotenoids in the Peel of Chihuahua Mango, *E3S Web Conf.*, vol. 38, no. 2026, pp. 3–5.
- [30] Faria-Silva, L., Gallon, C. Z., Purgatto, E., & Silva, D. M. (2017). Photochemical metabolism and fruit quality of Ubá mango tree exposed to combined light and heat stress in the field, *Acta Physiol. Plant.*, vol. 39, no. 10.
- [31] Zhu, H., Li, X., Zhai, W., Liu, Y., Gao, Q., Liu, J., & Zhu, Y. (2017). Effects of low light on photosynthetic properties, antioxidant enzyme activity, and anthocyanin accumulation in purple

- pak-choi (*Brassica campestris* ssp. *Chinensis* Makino), *PLoS One*, vol. 12, no. 6, pp. 1–17.
- [32] Vadiveloo, A., Moheimani, N. R., Cosgrove, J. J., Parlevliet, D., & Bahri, P. A. (2017). Effects of different light spectra on the growth, productivity and photosynthesis of two acclimated strains of *Nannochloropsis* sp., *J. Appl. Phycol.*, vol. 29, no. 4, pp. 1765–1774.
- [33] Ghasemzadeh, A., Jaafar, H. Z. E., Rahmat, A., Wahab, P. E. M., & Halim, M. R. A. (2010). Effect of different light intensities on total phenolics and flavonoids synthesis and anti-oxidant activities in young ginger varieties (*Zingiber officinale* Roscoe), *Int. J. Mol. Sci.*, vol. 11, no. 10, pp. 3885–3897.
- [34] Shui-Yuan, C., Xu, F., & Wang, Y. (2009). Advances in the study of flavonoids in *Ginkgo biloba* leaves, *J. Med. Plants Res.*, vol. 3, no. 13, pp. 1248–1252.
- [35] De-Ancos, B., Sánchez-Moreno, C., Zacarías, L., Rodrigo, M. J., Sáyago Ayerdí, S., Blancas Benítez, F. J., & González-Aguilar, G. A. (2018). Effects of two different drying methods (freeze-drying and hot air-drying) on the phenolic and carotenoid profile of ‘Ataulfo’ mango by-products, *J. Food Meas. Charact.*, vol. 0, no. 0, pp. 1–13.
- [36] Agatonovic-Kustrin, S., Kustrin, E., & Morton, D. W. (2018). Phenolic acids contribution to antioxidant activities and comparative assessment of phenolic content in mango pulp and peel, *South African J. Bot.*, vol. 116, pp. 158–163.
- [37] Pacheco-Ordaz, R., Antunes-Ricardo, M., Gutiérrez-Uribe, J. A., & González-Aguilar, G. A. (2018). Intestinal permeability and cellular antioxidant activity of phenolic compounds from mango (*Mangifera indica* cv. ataulfo) peels, *Int. J. Mol. Sci.*, vol. 19, no. 2.
- [38] Dorta, E., González, M., Lobo, M. G., Sánchez-Moreno, C., & de Ancos, B. (2014). Screening of phenolic compounds in by-product extracts from mangoes (*Mangifera indica* L.) by HPLC-ESI-QTOF-MS and multivariate analysis for use as a food ingredient, *Food Res. Int.*, vol. 57, pp. 51–60.
- [39] Salinas-Roca, B., Soliva-Fortuny, R., Welti-Chanes, J., & Martín-Belloso, O. (2018). Effect of pulsed light, edible coating, and dipping on the phenolic profile and antioxidant potential of fresh-cut mango, *J. Food Process. Preserv.*, vol. 42, no. 5, pp. 1–9.
- [40] Moreira-Rodríguez, M., Nair, V., Benavides, J., Cisneros-Zevallos, L., & Jacobo-Velázquez, D. A. (2017). UVA, UVB light, and methyl jasmonate, alone or combined, redirect the biosynthesis of glucosinolates, phenolics, carotenoids, and chlorophylls in broccoli sprouts, *Int. J. Mol. Sci.*, vol. 18, no. 11, pp. 1–20.
- [41] Surjadinata, B. B., Jacobo-Velázquez, D. A., & Cisneros-Zevallos, L. (2017). UVA, UVB and UVC light enhances the biosynthesis of phenolic antioxidants in fresh-cut carrot through a synergistic effect with wounding, *Molecules*, vol. 22, no. 4, pp. 1–13.
- [42] Wang, H., Gui, M., Tian, X., Xin, X., Wang, T., & li, J. (2017). Effects of UV-B on vitamin C, phenolics, flavonoids and their related enzyme activities in mung bean sprouts (*Vigna radiata*), *Int. J. Food Sci. Technol.*, vol. 52, no. 3, pp. 827–833.
- [43] Cruces, E., Flores-Molina, M. R., Díaz, M. J., Huovinen, P., & Gómez, I. (2017). Phenolics as photoprotective mechanism against combined action of UV radiation and temperature in the red alga *Gracilaria chilensis*, *J. Appl. Phycol.*, vol. 30, no. 2, pp. 1247–1257.
- [44] Pérez-Ambrocio, A., Guerrero-Beltrán, J. A., Aparicio-Fernández, X., Ávila-Sosa, R., Hernández-Carranza, P., Cid-Pérez, S., & Ochoa-Velasco, C. E. (2018). Effect of blue and ultraviolet-C light irradiation on bioactive compounds and antioxidant capacity of habanero pepper (*Capsicum chinense*) during refrigeration storage, *Postharvest Biol. Technol.*, vol. 135, no. August 2017, pp. 19–26.
- [45] Živanovic, B., Vidovic, M., Milic Komic, S., Jovanovic, L., Kolarz, P., Morina, F., & Veljovic Jovanovic, S. (2017). Contents of phenolics and carotenoids in tomato grown underpolytunnels with different UV-transmission rates, *Turkish J. Agric. For.*, vol. 41, pp. 113–120.
- [46] Jang, H. J., Lee, S. J., Kim, C. Y.,

- Hwang, J. T., Choi, J. H., Park, J. H., & Rho, M. C. (2017). Effect of Sunlight Radiation on the Growth and Chemical Constituents of *Salvia plebeia* R. Br., *Molecules*, vol. 22, no. 8, pp. 1–16.
- [47] Craver, J. K., Gerovac, J. R., Lopez, R. G., & Kopsell, D. A. (2017). Light Intensity and Light Quality from Sole-source Light-emitting Diodes Impact Phytochemical Concentrations within *Brassica* Microgreens, *J. Am. Soc. Hortic. Sci.*, vol. 142, no. 1, pp. 3–12.
- [48] U. Pérez-López *et al.*, Concentration of phenolic compounds is increased in lettuce grown under high light intensity and elevated CO<sub>2</sub>, *Plant Physiol. Biochem.*, vol. 123, pp. 233–241, 2018.
- [49] Kwaw, E., Ma, Y., Tchabo, W., Apaliya, M. T., Sackey, A. S., Wu, M., & Xiao, L. (2018). Impact of ultrasonication and pulsed light treatments on phenolics concentration and antioxidant activities of lactic-acid-fermented mulberry juice, *LWT - Food Sci. Technol.*, vol. 92, no. 2, pp. 61–66.